

# HOW RELIABLE ARE YOUR RESULTS?

## Reference Materials

Defined genomic DNA from microorganisms relevant for hygiene and contamination control.

# Genomic DNA Extracts



## Application

- Conventional and qPCR
- Specificity testing

These preparations contain genomic DNA extracted from low passage and defined microorganisms, prepared by phenol/chloroform extraction and subsequent column absorption methods. The DNA extract was partially sequenced to confirm identity. Titration was done by optical density measurement against a weight calf thymus DNA standard.

## Content

1 vial with genomic DNA extract with 10 ng +/- 2 ng, freeze-dried

1 vial with 200 µl 10 mM Tris-HCl buffer, pH 8.5, for dissolving the DNA

## Order information

Cat. No.	Species	Cat. No.	Species	Cat. No.	Species
51-0116	<i>Acholeplasma laidlawii</i>	51-1368	<i>Fluoribacter bozemanae</i> (syn. <i>Legionella bozemanae</i> )	51-0113	<i>Mycoplasma salivarium</i>
2127-30007	<i>Acinetobacter bamanii</i>			51-0124	<i>Mycoplasma synoviae</i>
2128-30187	<i>Aeromonas hydrophila</i>	2131-05934	<i>Geobacillus stearothermophilus</i>	2135-10036	<i>Neisseria meningitidis</i>
2101-00819	<i>Aspergillus fumigatus</i>			2136-13387	<i>Proteus vulgaris</i>
51-0031	<i>Bacillus cereus</i>	2132-30104	<i>Klebsiella pneumoniae</i>	51-0071	<i>Pseudomonas aeruginosa</i>
51-0010	<i>Bacillus subtilis</i>	2133-70603	<i>Klebsiella pneumoniae</i> , ESBL+	2116-04479	<i>Proteus mirabilis</i>
2129-02046	<i>Bacillus thuringiensis</i>			51-7058	<i>Salmonella enterica</i>
51-3415	<i>Bordetella parapertussis</i>	51-1723	<i>Lactobacillus acidophilus</i>	2117-30121	<i>Serratia marcescens</i>
51-5571	<i>Bordetella pertussis</i>	51-1370	<i>Legionella dumofii</i>	2137-04782	<i>Shigella flexneri</i>
2130-07288	<i>Burkholderia capacia</i>	51-1533	<i>Legionella jordanis</i>	2138-05570	<i>Shigella sonnei</i>
2102-04688	<i>Campylobacter jejuni</i>	51-0101	<i>Legionella pneumophila</i>	51-0164	<i>Spiroplasma citri</i>
51-1386	<i>Candida albicans</i>	51-1514	<i>Legionella pneumophila</i> subsp. <i>fraseri</i>	51-0231	<i>Staphylococcus aureus</i>
2103-11226	<i>Candida glabrata</i>			51-0044	<i>Staphylococcus epidermidis</i>
2104-11947	<i>Candida guilliermondii</i>	51-1515	<i>Legionella pneumophila</i> subsp. <i>pascullei</i>	2118-20328	<i>Staphylococcus hominis</i>
2105-70624	<i>Candida haemulonii</i>			2119-20263	<i>Staphylococcus haemolyticus</i>
2106-30039	<i>Citrobacter freundii</i>	51-3361	Methicilin-resistant <i>Staphylococcus aureus</i> (MRSA)	2122-20480	<i>Streptococcus bovis</i>
2125-90874	<i>Candida tropicalis</i>			2123-06176	<i>Streptococcus dysgalactiae</i>
2107-04595	<i>Citrobacter koseri</i>	2134-30164	<i>Morganella morganii</i>	2126-20523	<i>Streptococcus mutans</i>
51-0792	<i>Clostridium acetobutylicum</i>	51-0030	<i>Micrococcus luteus</i>	51-0566	<i>Streptococcus pneumoniae</i>
2108-00756	<i>Clostridium perfringens</i>	51-0129	<i>Mycoplasma arginini</i>	2139-20068	<i>Streptococcus sanguinis</i>
51-0053	<i>Enterobacter aerogenes</i>	51-0162	<i>Mycoplasma arthritidis</i>	2120-20229	<i>Staphylococcus saprophyticus</i>
2110-30054	<i>Enterobacter cloacae</i>	51-0117	<i>Mycoplasma fermentans</i>		
2111-20680	<i>Enterococcus casseliflavus</i>	51-0115	<i>Mycoplasma gallisepticum</i>	2121-50170	<i>Stenotrophomonas maltophilia</i>
2112-30633	<i>Enterococcus durans</i>	51-0195	<i>Mycoplasma genitalium</i>		
51-0478	<i>Enterococcus faecalis</i>	51-0111	<i>Mycoplasma hominis</i>	51-0177	<i>Ureaplasma urealyticum</i>
2113-20477	<i>Enterococcus faecium</i>	51-0130	<i>Mycoplasma hyorhinis</i>	2140-04780	<i>Yersinia enterocolitica</i>
2114-20160	<i>Enterococcus hirse</i>	51-0112	<i>Mycoplasma orale</i>	2141-08992	<i>Yersinia pseudotuberculosis</i>
51-0083	<i>Escherichia coli</i>	51-1746	<i>Mycoplasma penetrans</i>		
2115-08579	<i>Escherichia coli</i> O157:H7	51-0119	<i>Mycoplasma pneumoniae</i>		

# PCR Quantification Standards



## Application

- Conventional and qPCR
- Standard curves, evaluation of assay performance
- Preparation of dilution series for quantification
- Low titer (e.g.  $3 \times \text{LOD}_{95}$ ) controls

Standardization and quantification of nucleic acid detection is a difficult task as there are no reliable standards for low nucleic acid copy numbers. Detection methods such as PCR and NASBA are prone to inhibition caused by many substances commonly found in plant and animal tissues, food matrices and extraction solutions. When co-purified with DNA or RNA, inhibitors reduce amplification efficiency, causing an underestimation of the quantity of target nucleic acid or even false-negative results.

Minerva Biolabs calibration reagents contain genomic DNA extracted from low passage and defined microorganisms. The DNA is manufactured by means of phenol/chloroform extraction with ethanol precipitation and subsequent column absorption methods. The DNA extract was partially sequenced and the sequence aligned to confirm identity. Titration was done after photometric quantification of the preparation standard and dsDNA fluorometric quantification against a synthetic standard.

For the first time, qPCR user are able to include a precise low count of DNA copies in their assays, providing them with a true value for the estimation of detection limits and the comparison of different detection methods.

## Content

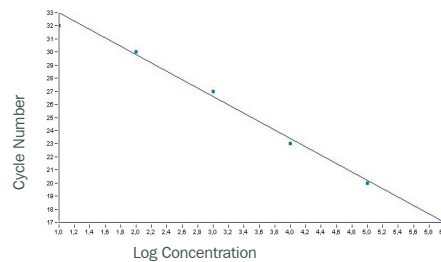
1 vial with DNA,  $1 \times 10^8$  genomes, freeze-dried

3 vials with 2 ml of Tris-HCl buffer, 10 mM, pH 8.5., for dissolving the DNA and preparing dilutions

## Order information

Cat. No.	Species
52-0116	<i>Acholeplasma laidlawii</i>
52-5571	<i>Bordetella pertussis</i>
52-0083	<i>Escherichia coli</i>
52-0101	<i>Legionella pneumophila</i>
52-0129	<i>Mycoplasma arginini</i>
52-0117	<i>Mycoplasma fermentans</i>
52-0115	<i>Mycoplasma gallisepticum</i>
52-0130	<i>Mycoplasma hyorhinis</i>
52-0112	<i>Mycoplasma orale</i>
52-0119	<i>Mycoplasma pneumoniae</i>
52-0124	<i>Mycoplasma synoviae</i>
52-0164	<i>Spiroplasma citri</i>
52-0071	<i>Pseudomonas aeruginosa</i>

Standard Curve



Real-time Amplification Plot

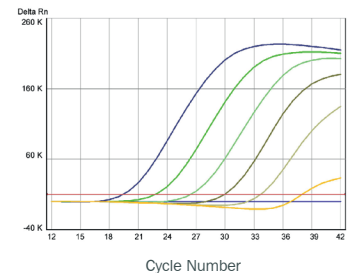


Fig. Quantification of *Mycoplasma pneumoniae* DNA. Logarithmic plot of fluorescence vs cycle number (Venor®GeM qER, platform: ABI Prism® 7500). Template DNA ranging from  $2 \times 10^5$  - 2 genome equivalents.

# 10CFU™ Sensitivity Standards

For validating robustness and detection limit of molecular mycoplasma test methods in presence of the sample matrix.



## Application

European Pharmacopoeia 2.6.7 “Mycoplasma” requires a sensitivity of 10 CFU/ml sample volume for NAT-based methods like PCR to replace the traditional culture method. This feature of the test method must be shown by the performing lab as part of the robustness testing in presence of the sample matrix. As most cell culture labs and production facilities cannot accept vital mycoplasma in their facility or do not have access to a microbiology lab able to cultivate mycoplasma, these preparations allow safe and reliable validation of the procedure.

The mycoplasma have been cultivated in culture broth described in EP 2.6.7, titrated immediately in culture broth and plated for quantification in colony forming units (CFU/ml). Each dilution series has been performed in multiple by different operators for highest precision. The mycoplasma broth was harvested in the early logarithmic phase of the growth to avoid a high ratio of dead mycoplasma particles and correspondingly a high GU\*:CFU ratio. All strains have been obtained from official culture collections and cultivated in low passages.

Each vial contains 10 CFU of inactivated mycoplasma. By adding the sample matrix of interest a sample according to EP 2.6.7 is prepared which has to be tested positive by the method applied. Obviously, the inactivated sample material is not suitable for the culture method anymore. As a result of proficiency tests on DNA amplification methods for mycoplasma detection it became obvious that in means of highest sensitivity DNA extraction is indispensable. The extract can directly be used for PCR.

\* Please note: This standard material was not titrated for genome copies (GU) as EP 2.6.7 does not provide sensitivity limits on DNA level. No guarantee for a particular GU:CFU ratio is provided with this product and the ratio may vary from lot to lot.

## Package Content

3 vials with 10 CFU of the corresponding mycoplasma species  
2 negative control vials  
For the mycoplasma set: 2 vials with 10 CFU of each mycoplasma species listed in the EP 2.6.7 (18 vials in total)  
2 negative controls

## Catalogue Number

102-1003	<i>Mycoplasma arginini</i>	102-6003	<i>Mycoplasma fermentans</i>
102-2003	<i>Mycoplasma orale</i>	102-7003	<i>Mycoplasma hyorhinis</i>
102-3003	<i>Mycoplasma gallisepticum</i>	102-8003	<i>Acholeplasma laidlawii</i>
102-4003	<i>Mycoplasma pneumoniae</i>	102-9003	<i>Spiroplasma citri</i>
102-5003	<i>Mycoplasma synoviae</i>	102-0002	Mycoplasma Set

Minerva Biolabs GmbH

Koepenicker Strasse 325 . D-12555 Berlin . Germany  
Tel. +49 (0)30 2000 437-0 . Fax +49 (0)30 2000 437-9  
info@minerva-biolabs.com . www.minerva-biolabs.com

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